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Patterned cell culture substrates created by hot embossing of tissue culture treated polystyrene

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Abstract

Patterning materials such that they elicit a different cell response in different regions would have significant implications in fields such as implantable biomaterials, in vitro cell culture and tissue engineering and regenerative medicine. Moreover, the ability to pattern polymers using inexpensive, currently available processes, without the need for adding proteins or other biochemical agents could lead to new opportunities in biomaterials research. The research reported here demonstrates that by combining the plasma surface treatments used to create commercial grade tissue culture treated polystyrene, with controlled hot embossing processes, that distinct regions can be created on a substrate that result in spatial control of endothelial cell adhesion and proliferation. As well as the topographical changes that result from hot embossing, significant changes in surface chemistry and wettability have been observed and characterised and the resultant effects on endothelial cell responses evaluated. By spatially controlling endothelial cell adhesion, proliferation and subsequent angiogenesis, the processes outlined here have the potential to be used to create a range of different substrates, with applications in the development

of assays for high throughput screening, the patterning of implantable biomaterials or the development of smart scaffolds for tissue engineering.

1. Introduction

The ability of endothelial cells to form a network of cord and tube structures forms the basis of both research into angiogenesis and the provision of a vascular network for 3-dimensional tissue engineered constructs. Gradients of growth factors [1] and the containment of endothelial cells either within channels [2] or around fibre structures [3] have been used to spatially control the culture of endothelial cells and thus the subsequent processes of angiogenesis. Spatial control of angiogenesis could have applications in the development of new angiogenesis assays, as well as in the development of strategies for vascularisation of tissue engineered constructs.

Endothelial cell responses can also be altered by culturing on topographically and/or chemically modified surfaces. For example, the alignment of endothelial cells to micro- and nano-scale grooves in polydimethylsiloxane (PDMS), coated with a fibronectin monolayer [4], and to nano- and micro-scale grooves in polyurethane substrates, especially those with groove widths greater than 800 nm, but not to arrays of pore structures [5] have been observed. Other studies have shown that alignment is more pronounced for deeper groove depths on PDMS substrates [6, 7] and also with smaller groove widths [7]. The shape of endothelial cells has also been modified by culturing on cell-adhesive and non-adhesive regions that are created on substrates at the micro-scale. The resulting “shape-engineered” endothelial cells are elongated and aligned, and also show alignment of the actin cytoskeleton [8]. In addition, cells aligned in this way show a reduction in mRNA expression for vascular cell adhesion molecule-1 (VCAM-1), a result which is similar to that found with endothelial cells aligned by exposure to shear stress [9]. On lines of collagen or fibronectin, endothelial cells and the actin cytoskeleton will align parallel to the lanes [10].

It is clear therefore, that spatial control of where endothelial cells are cultured, as well as control of cell shape can both play a role in the responses of endothelial cells to an engineered surface. Modifications in surface topography and surface chemistry either individually, or in combination, can provide for a means of both spatial control and shape control, and have the potential to elicit specific cell responses, including changes in cell adhesion,

differentiation and alignment. Significant differences in cell response can be observed for relatively subtle changes in the biomaterial surface on which the cells are cultured, and by using micropatterning techniques it is possible to create adjoining regions where cell responses are different. Many of these patterning methods involve the use of proteins or biochemical factors, with resultant shelf-life and refrigerated storage requirements.

Polystyrene has been used for the culture of adherent cells for many years, as it is easy to process, and is inexpensive. As a result, these surfaces are well understood and are often used as control surfaces in cell studies. Native polystyrene, however is not a suitable surface for adherent cell culture [11, 12] and needs to be pre-treated in order to improve the wettability of the surface and thus improve cell adhesion. These pre-treatments typically both increase the surface roughness and add oxygenated chemical groups to the surface [13, 14], producing tissue culture treated polystyrene (TCPS).

This research uses hot embossing to make specifically defined changes to the topography and surface chemistry of TCPS substrates, and by characterising the changes in topography, roughness and surface chemistry, this study shows that spatial control over adhesion, proliferation, alignment and subsequent processes of angiogenesis can be affected by the TCPS surface, and allows for a simple method of producing adhesive and non-adhesive regions on TCPS substrates.

2. Materials and methods

2.1. Hot embossed micro-featured substrates

Tissue culture treated polystyrene (TCPS) substrates were prepared from commercial grade T-175 tissue culture flasks (Sarstedt, Germany). 100 mm diameter discs were cut from the cell culture surface of the flasks using a Thermocut hot wire cutter (Proxxon, Germany). Untreated polystyrene (PS) discs were cut from 1.2 mm thick PS sheets (Goodfellow, UK) as a control.

Samples were hot embossed at 110 °C and 10 kN embossing force under vacuum (<1 mbar) in an EVG520 HE automated hot-embossing tool (EVGroup, Austria) using silicon stamps, fabricated using photolithography and plasma etch processes at Tyndall National Institute (Cork, Ireland) as part of the National Access Programme. A number of feature arrays were produced by this hot embossing process and these are described in Table 1.

Table 1

Names and descriptions of micro-features created by hot embossing

Pattern name	Feature type	Feature width (μm)	Feature height (μm)	Array details
3.2 μm ridge	Rectangular ridge	3.2	1	10 mm long parallel ridges with 3.2 μm spacing
3.2 μm pillar	Circular pillar	3.2	1	Hexagonal close packed array with 4.8 μm centre-to-centre spacing
3.2 μm short ridge	Rectangular ridge	3.2	1	200 mm long parallel ridge array with 3.2 μm spacing and separated by 200 μm from the adjacent array

3.2 μm ridge arrays were chosen based on the ability to fabricate hot embossing stamps using standard contact photolithography and being in the size range that has been shown to result in alignment and elongation of endothelial cells. The pillar arrays have the same proportion of raised and planar areas as the ridges, but do not have the specific orientation of the ridge arrays. The short ridge arrays were investigated to see the effect of patterned and unpatterned regions in close proximity.

2.2. Surface characterisation

2.2.1. Stylus profilometry

A Dektak 8 Advanced Development Profiler (Veeco, USA) was used to measure feature dimensions produced by the hot embossing process and to measure surface roughness of the polymer surfaces after embossing. The profiler was fitted with a 5 μm diameter stylus at a scan force of 3 mg. Line scans of 1000 μm were performed on samples with this stylus to investigate step heights.

2.2.2. Static contact angle measurement

The wettability was measured using a CAM 200 (KSV Instruments, UK) on the various embossed and non-embossed substrates. The static water contact angle was measured using the sessile drop method using a 5 μL drop of DI water.

2.2.3. Atomic force microscopy (AFM)

As well as measuring the roughness using stylus profilometry, surface roughness was also measured on a smaller number of samples by atomic force microscopy (AFM) using a Dimension 3100 Atomic Force Microscope (Veeco Instruments Inc., USA) fitted with a TESPA tip (Veeco Instruments Inc., USA) with resonant frequency 246–375 kHz. $10\text{ }\mu\text{m} \times 10\text{ }\mu\text{m}$ scans were performed at 1 Hz scanning rate resulting in 512×512 resolution maps. The Amplitude set point was 1.46 V. After scanning, results were analysed using Nanoscope v6.14R1 software (Veeco Instruments Inc.), where the arithmetic mean roughness R_a was calculated for the surface and a 3-D rendering of the surface was produced.

2.2.4. X-ray photoelectron spectroscopy (XPS)

X-ray photoelectron spectroscopy (XPS) was used to determine the chemical composition of the PS and TCPS before and after the hot embossing process. The technique was used to quantify the carbon and oxygen concentration at the polymer surface and to therefore evaluate the effect of processing conditions on the oxygen containing functional groups that are present on TCPS after tissue culture treatment.

XPS analysis was performed using a Axis Ultra DLD spectrometer (Kratos Analytical, Japan) using monochromated Al $K\alpha$ X-rays operating at 15 kV and 10 mA. The operating pressure was lower than 6×10^{-6} Pa. A hybrid lens mode was used during analysis. Charge neutralisation was achieved using an immersion lens with a filament current of between 1.9 and 2.1 mA at a charge balance voltage of between 3.2 and 3.6 V. Three spots were analysed per sample and wide energy survey scans (0–1300 eV binding energy) as well as high resolution spectra for C 1s (278–295 eV) and O 1s (525–540 eV). Pass-energy was 160 eV for the wide energy survey scans and 20 eV for the high resolution spectra.

Quantification of the atomic % oxygen at the surface from the high resolution spectra was conducted by subtracting a linear background and calculating the area under the peaks on the C 1s and O1s spectra using Vision 2.2.8 software (Kratos Analytical, Japan). At least two samples of each type were analysed.

2.3. Cell culture

10 mm square coupons were used for cell culture tests. Bovine Aortic

Endothelial Cells (BAEC) were seeded onto pristine and hot embossed TCPS coupons at 5×10^4 cells/cm² and cultured in Minimum Essential Media, with the addition of 10 % Foetal Bovine Serum and antibiotic and antimycotic supplements at 37 °C and 5 % CO₂ in a cell culture incubator. Primary cells between passage 4 and 8 were used in the cell culture experiments described here. Cell numbers, aspect ratio and alignment were assessed after 48 and 72 h and the longer term effects were assessed after a further 7–10 days in culture.

2.3.1. Fluorescent and immunofluorescent labelling

In order to visualise morphological and cytoskeletal changes in response to the micro-features, samples were stained using both fluorescent and immunocytochemical labelling procedures. Cell monolayers were fixed in a solution of 4 % paraformaldehyde containing 0.1 % Triton X-100 if the plasma membrane was to be permeabilised. As required, either the plasma membrane was labelled using Alexa-Fluor 546 conjugated Wheatgerm Agglutinin or the actin cytoskeleton was labelled using Alexa-Fluor 488 conjugated Phalloidin. For immunocytochemical labelling, samples were incubated in a primary antibody, targeted against β -tubulin, for 1 h at 37 °C, followed by incubation with a fluorescent secondary antibody conjugated with Alexa Fluor 594, targeted against the primary antibody, for 45 min at 37 °C. Cell nuclei were labelled using 4',6-diamidino-2-phenylindole (DAPI) (Molecular Probes, USA). After staining, cell monolayers were mounted using Vectashield mounting media, covered with a glass cover slip and sealed with clear varnish. Mounted samples were stored at 4 °C in the dark prior to imaging.

2.3.2. Confocal laser scanning microscopy (CLSM)

Labelled cells were imaged using confocal laser scanning microscopy (CLSM) using a LSM Pascal exciter (Zeiss, Germany) mounted on a Zeiss Axioimager M1 microscope (Zeiss, Germany) fitted with a 488 nm Argon laser, A 405 nm diode laser and a 543 nm Helium–Neon laser. Typically, as most samples were either dual or triple labelled, a multi-track setup was used and an image stack was then captured at the required resolution and z-step height. Once the z-stack had been captured, this was processed into a 2D image using the LSM Image Examiner software (Zeiss, Germany).

2.3.3. Scanning Electron Microscopy (SEM)

Samples were fixed and prepared for Scanning Electron Microscopy (SEM)

using the following protocol. After gentle washing in PBS, primary fixation was performed using 2.5 % glutaraldehyde in phosphate buffered saline (PBS) at pH 7 for 20 min at room temperature. After washing several times with PBS, secondary fixation was performed in 1 % osmium tetroxide solution for 1 h. The samples were again washed several times in PBS before being dehydrated using a series of graded alcohol washes. After ethanol dehydration, final chemical dehydration was carried out using hexamethyldisilazane (HMDS) before being air dried overnight.

Once dehydrated, the samples for SEM were coated in a conductive coating of gold in a K500X sputter coater (Emitech, USA) and analysed in a Quanta 200 3D scanning electron microscope (FEI, USA).

2.3.4. Image analysis

Fluorescent micrographs were analysed using Image J [15]. Colour or greyscale images were converted to an 8-bit greyscale type, before converting to a binary image by applying upper and lower thresholds. The threshold levels were manually adjusted to obtain the best representation of the cells or cell nuclei. Once the binary image was obtained, image analysis was performed using the particle analyser in Image J.

For cell counting, the particle size range considered was adjusted to ensure that image artefacts that were smaller than the cell nuclei were not counted. In order to confirm the accuracy of this methodology, the nuclei on at least two images per sample were counted manually and compared with the results obtained by image analysis. Particle size range was adjusted so that these counts were within 5 % of each other for all the images considered. From the cell numbers, the cell density was then determined from the full frame areas, based on the microscope objective used. Direct cell counting, based on microscopy and image analysis, was chosen as the most appropriate method for quantifying adhesion and proliferation on the various substrates, as it was possible to count cell numbers specifically for each different surface type, when a range of different surfaces were present on some samples. As the purpose was to determine the cell response to the different surface features, it was important to be able to consider these surfaces individually, which would not have been possible using biochemical assays.

To determine cell alignment and aspect ratio, fluorescent images were processed using ImageJ using a similar methodology to that employed by

O’Cearbhaill et al. [16]. Within the particle analyser, an ellipse was fitted to each cell. From the fitted ellipses, the lengths of the major and minor axes were measured, as well as the angle the major axis made with the long axis of the image frame. From the axis lengths, the aspect ratio, sometimes known as the cell elongation index [4, 17] was calculated by dividing the major axis length by the minor axis length. An aspect ratio index of 1 denotes a cell that is as wide as it is long, while higher numbers indicate more elongated cells. Aspect ratios were measured on all cells from at least eight images per sample, and the mean aspect ratio calculated for each set of experimental conditions. *t*-tests, assuming equal variances were used to assess the statistical significance of any differences observed.

In order to determine the alignment of cells, histograms of the major axis angle were plotted for each sample using a method similar to Punchard et al. [18] and Charest et al. [19]. If the cells were not preferentially aligned in any direction, then the histogram would show a uniform distribution. If, however, there was preferential alignment, then a larger proportion of cells would be seen with their major axes in this range.

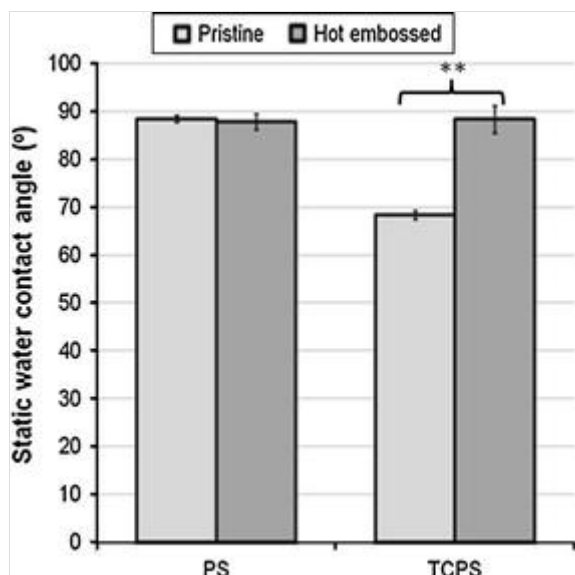
3. Results

3.1. Micro-features created by hot embossing

Scanning Electron and optical micrographs of the micro-features produced by hot embossing are shown in Fig. 1. The features are accurately and repeatably replicated across the 10 mm square array. The step heights of the micro-features were measured by stylus profilometry, and a sample scan of a test region can be seen alongside the micrographs in Fig. 1.

Fig. 1

SEM images of hot embossed micro-features in TCPS showing **a** 3.2 μm ridges and **b** 3.2 μm pillars. **c** shows a stylus profilometer scan indicating the 1 μm step height of the features. Image **d** is an optical micrograph showing the short ridge array adjacent to an unfeatured region

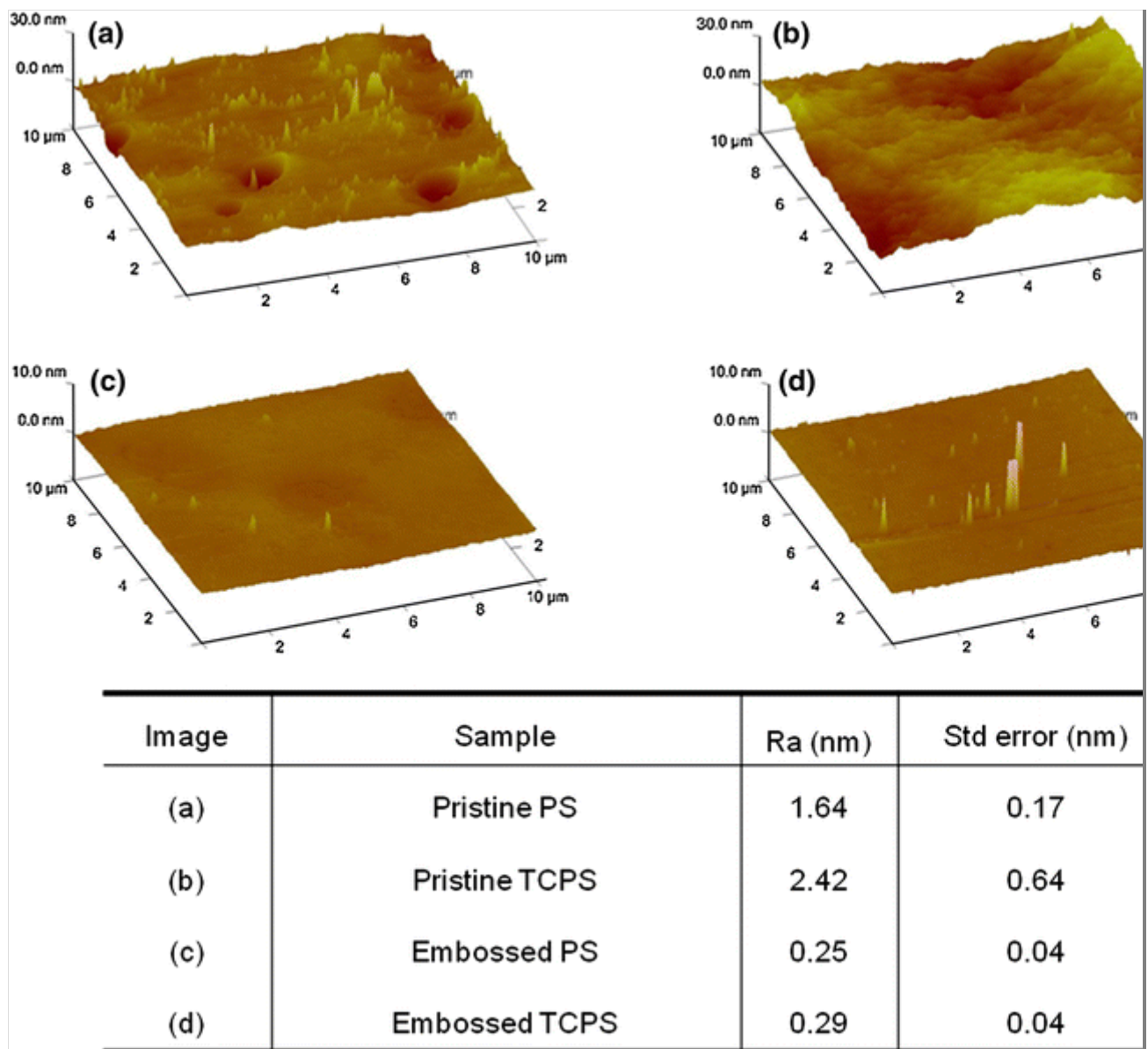


3.2.2. Atomic force microscopy (AFM)

AFM analysis shows that the surface of both the PS and TCPS substrates is significantly less rough after hot embossing. Images and [Ra values](#) are shown in [Fig. 3](#) and the [Ra values are shown in Table 2](#). Polished silicon is very smooth, with a typical Ra value of lower than 1 nm, so it is not surprising that the hot embossing process creates a very smooth polymer surface.

Fig. 3

3-D rendering of $10\ \mu\text{m} \times 10\ \mu\text{m}$ AFM scans and arithmetic mean surface roughness on **a** pristine PS, **b** pristine TCPS, **c** embossed PS and **d** embossed TCPS, showing the significant reduction in surface roughness after hot embossing. *Note* that the z-scale for the embossed samples is $\pm 10\ \text{nm}$ while that for the pristine surfaces is $\pm 30\ \text{nm}$

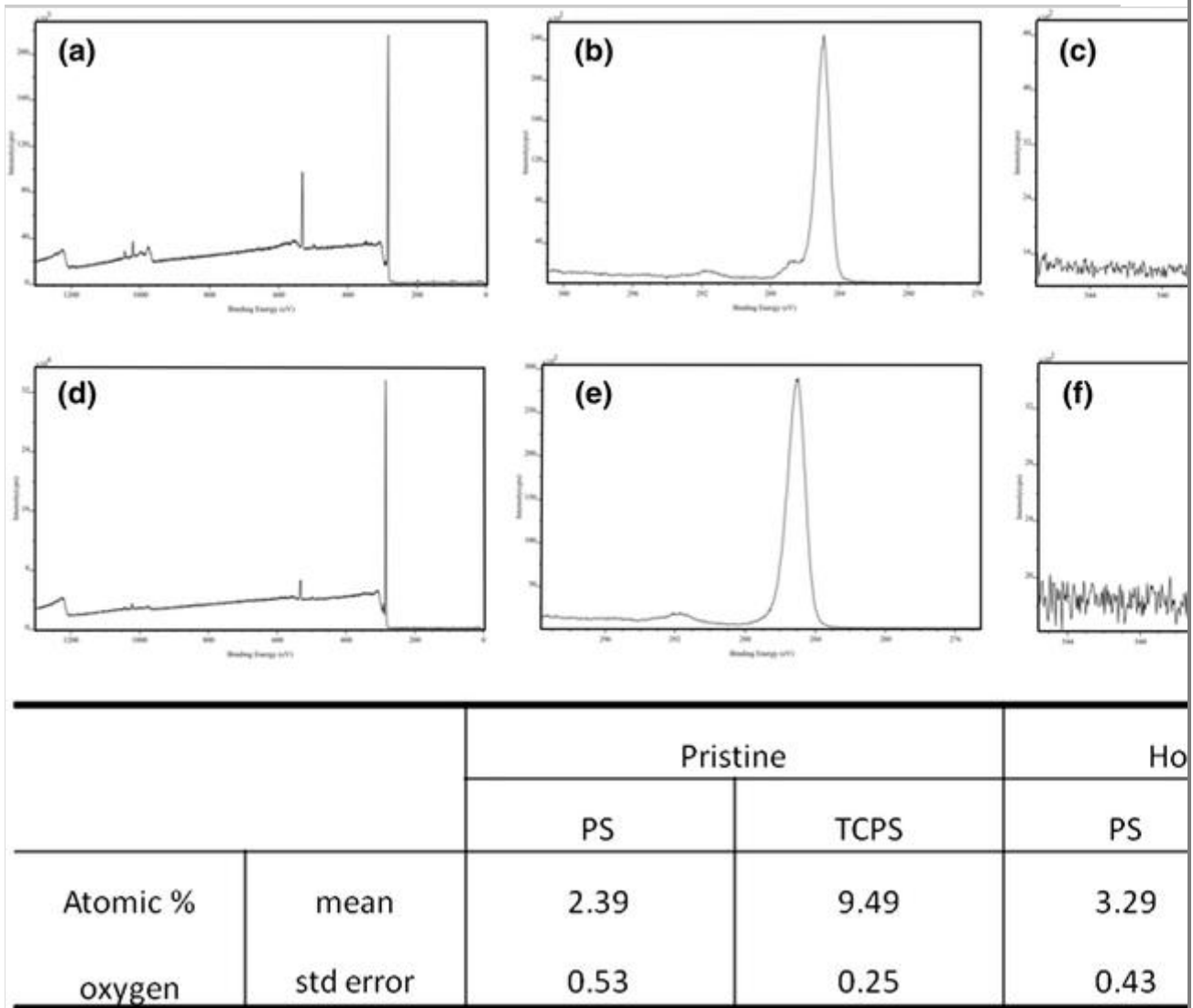


3.2.3. X-Ray photoelectron spectroscopy (XPS)

Chemical changes to the TCPS surface were evaluated using XPS. The spectra in Fig. 4 indicate that there is a significant reduction in the atomic % oxygen at the surface of the TCPS substrate after hot embossing. The quantification of the area under the oxygen peaks is shown in Fig. 4. The value of 9.49 % for TCPS is typical of the atomic % oxygen found after tissue culture treatment.

Fig. 4

XPS spectra for pristine TCPS showing **a** wide energy survey scan, **b** high resolution carbon peak, **c** high resolution oxygen peak and for hot embossed TCPS showing **d** wide energy survey scan, **e** high resolution carbon peak, **f** high resolution oxygen peak. Quantification data are shown for pristine and hot embossed PS and TCPS substrates showing a reduction in atomic oxygen after hot embossing for TCPS. The Pristine TCPS surface is statistically significantly different from all of the other surfaces at

$P < 0.01$


3.3. Cell culture

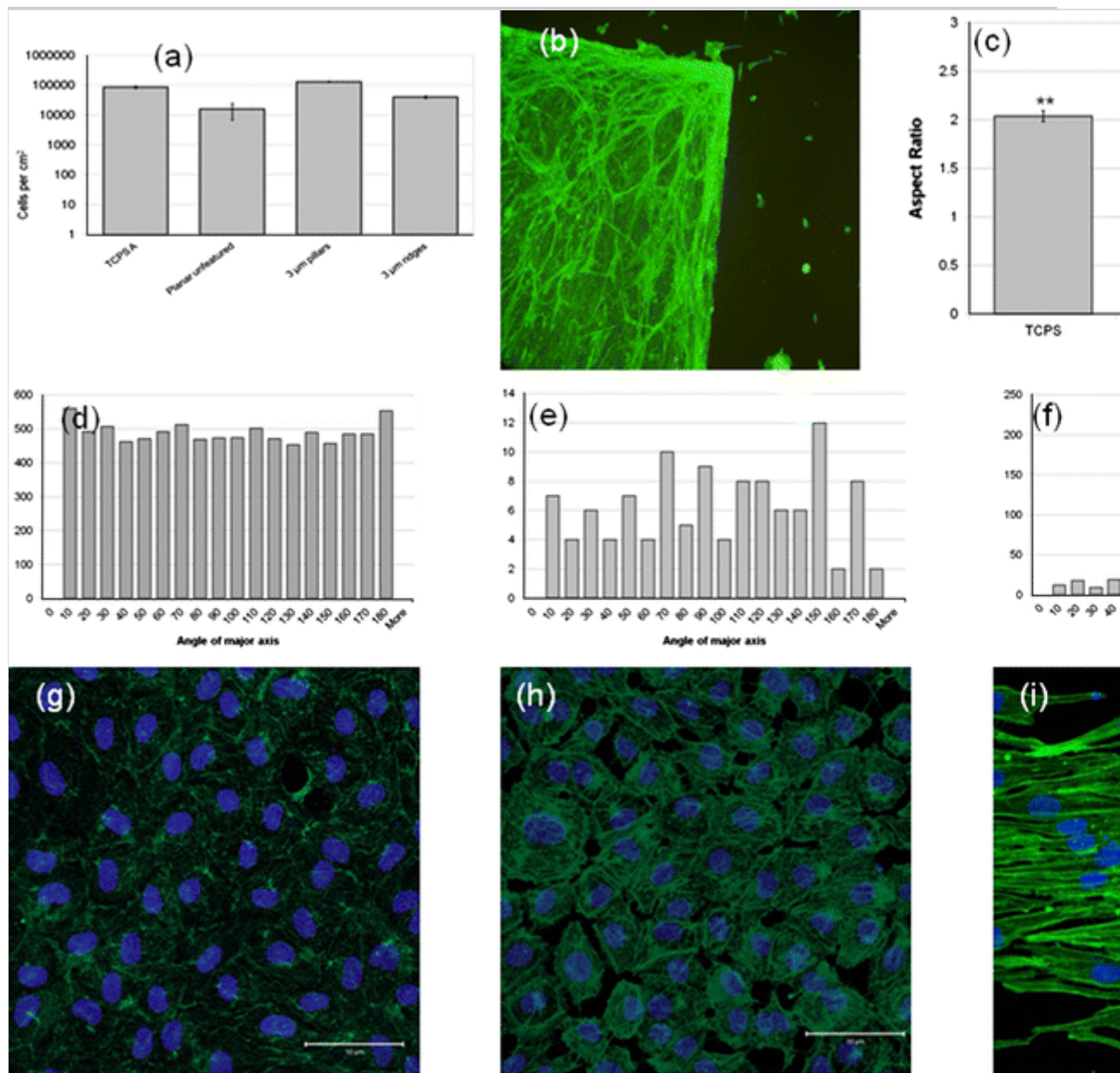
3.3.1. Quantification of cell numbers

After 48 and 72 h in culture, cell numbers were evaluated using fluorescent microscopy and image analysis, with results after 72 h shown in Fig. 5. After 48 h, cell monolayers were not yet confluent on any of the substrates, while after 72 h the cells were confluent on the 3.2 μm pillars and ridges as well as the pristine TCPS. When confluent, the cell density was lower on the 3.2 μm ridges than on the 3.2 μm pillars or the pristine TCPS.

Fig. 5

a cell numbers after 72 h in culture, showing almost an order of magnitude difference in cell number between pristine TCPS and planar embossed TCPS. **b** BAECs cultured on 3.2 μm pillars showing preferential adhesion and proliferation on the micro-featured areas, and very little adhesion to the planar unfeatured areas. **c** shows the aspect ratio

of BAECs cultured on different substrates, preferential alignment histograms after 72 h in culture for **d** non-aligned BAECs on pristine TCPS, **e** non-aligned BAECs cultured on 3.2 μm pillars and **f** aligned BAECs cultured on 3.2 μm ridges. CLSM images of BAECs showing actin (green) and nuclei (blue) cultured on **g** pristine TCPS, **h** 3.2 μm pillars and **i** 3.2 μm ridges after 72 h in culture



The planar embossed TCPS substrates showed the lowest cell numbers, as shown in Fig. 5a. It can be clearly seen in the micrographs in Fig. 5b, that where micro-features and planar embossed surfaces exist side-by-side, that BAECs preferentially adhere and proliferate on the micro-features.

3.3.2. Cell aspect ratio and alignment

As is expected, cells align and elongate in the direction of the micro-scale

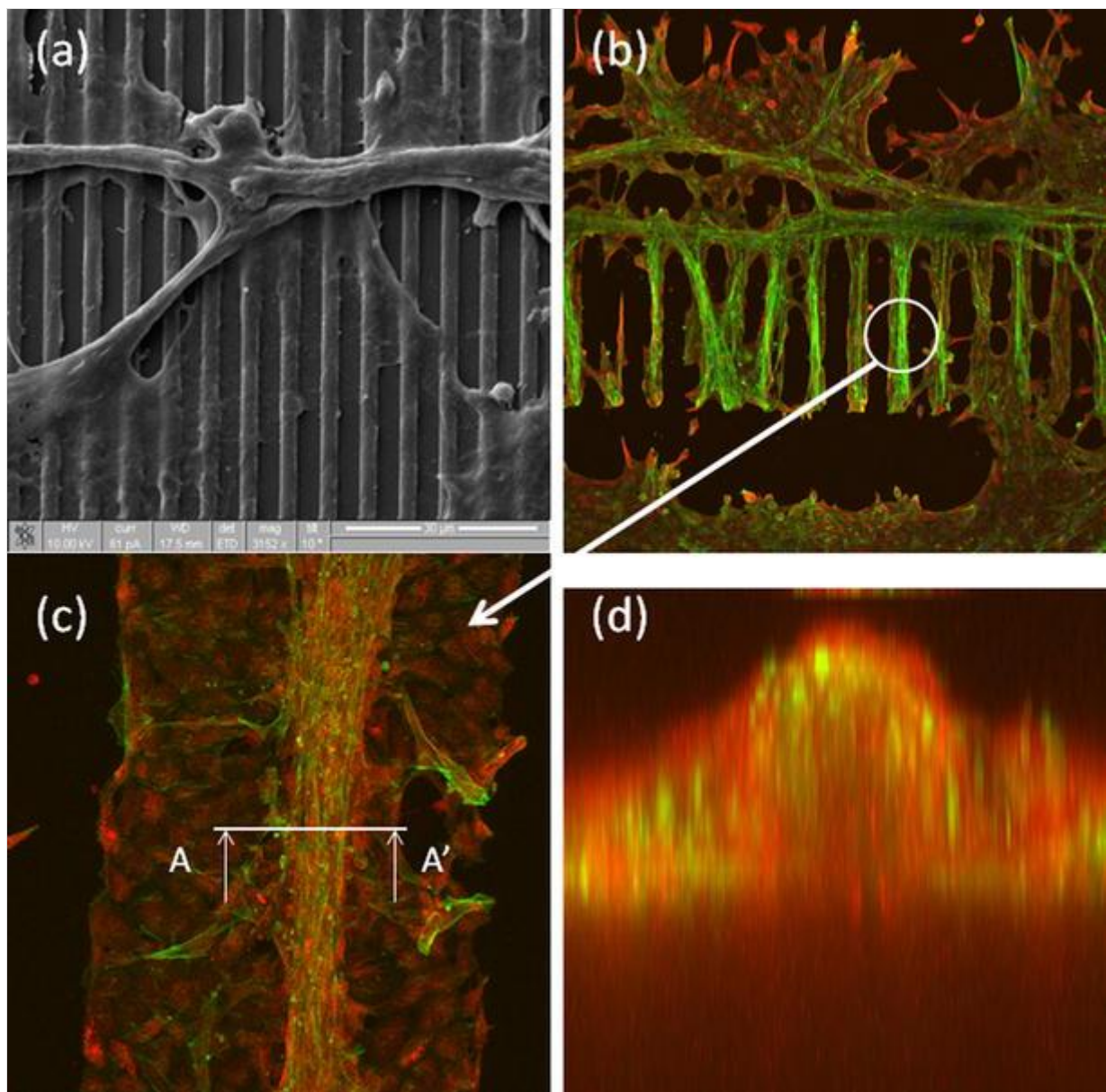
ridge features. The aspect ratios of BAECs after 48 h in culture are shown in Fig. 5c, and the histograms showing the preferential alignment of BAECs on the ridge features are shown in Fig. 5d, e. No significant difference is observed in the aspect ratio or alignment of cells cultured on any of the pillar features or the pristine TCPS. Representative CLSM images of BAECs on microfeatured surfaces are shown in Fig. 5f, g.

3.3.3. Directional tubule formation

After a further 7–10 days in culture (10–13 days in total), BAECs appear to have formed cord or tube structures on the substrates with the 200 μm long ridges. SEM and CLSM images of these tubules are shown in Fig. 6. These cords or tubes are formed in a direction that is perpendicular to the individual ridge features, i.e. they are formed parallel to the long axis of the patterned strip. The diameter of these structures varies, suggesting that they may initially form as cords or capillaries and subsequently grow in diameter.

Fig. 6

Tubules formed on short ridge arrays after 10 days in culture: **a** shows a scanning electron micrograph of tubules, **b** is a low magnification CLSM image of a number of tubules formed on several parallel short ridge arrays. Image **c** is a projection of a CLSM image stack of a single tubule on a short ridge, similar to the circled region in (**b**). Image (**d**) is a projected image from the CLSM stack of the cross section A–A', showing a tubular structure



SEM and CLSM images of these tubules are shown in Fig. 6.

4. Discussion

4.1. The effect of hot embossing on tissue culture treated polystyrene

Hot embossing has been used here to produce repeatable micro-scale features in both PS and TCPS substrates. Hot embossing of PS shows a reduction in surface roughness after hot embossing using a silicon stamp, as measured by AFM, but no significant change in water contact angle or in the surface chemistry of PS, as measured by XPS. In contrast, when tissue culture treated polystyrene (TCPS) is considered, the surface analysis undertaken here indicates that the hot embossing process changes the chemistry of the surface significantly as well as changing its topography. The contact angle of the

TCPS increases significantly after embossing. This research has shown that embossing TCPS substrates at 110 °C and 10 kN produces a surface that is significantly more hydrophobic than pristine TCPS.

Polished silicon typically has a surface roughness of 1 nm or less [20], measured by AFM, so it is to be expected that embossing PS or TCPS with such a surface will result in a reduction in surface roughness.

The effects of hot embossing on TCPS are supported by the XPS analysis which indicates that atomic oxygen concentration at the surface for the pristine TCPS substrate shows 9.5 at.% oxygen at the surface, while the pristine PS substrate contains approximately 2.4 at.% oxygen at the surface. These values are comparable to those found in the literature for polystyrene and plasma treated polystyrene [11]. The surface treatment of polymers using plasmas has been shown to be degraded by ageing, where the mobility of the polymer chains result in a gradual movement of polar functional groups into the bulk of the polymer [21]. This mobility will be greatly increased by heating the polymer to its glass transition temperature during the hot embossing process, consistent with the results observed here. The combined effect of a reduction in surface roughness and a reduction in the oxygen containing polar functional groups at the polymer surface is consistent with the increase in water contact angle observed in this research.

The effect of hot embossing on the TCPS surface chemistry has not been reported previously, and has important implications for the subsequent adhesion of cells to these surfaces in the context of the use of hot embossing processes for the production of micro-scale features in treated polymer surfaces for cell culture, as the maximum cell adhesion of a range of cell types cells has been reported at water contact angles of around 55° [13, 22], with poorer adhesion at both higher and lower contact angles. It is therefore expected that cells cultured on TCPS and hot embossed TCPS will behave differently. Considering the low levels of cell adhesion of to pristine PS, adhesion of cells to embossed, planar TCPS surfaces is likely to be lower than to non-embossed TCPS surfaces.

4.2. BAEC response to hot embossed TCPS surfaces

4.2.1. Cell numbers, aspect ratio and alignment

BAECs adhere significantly less well to planar, non-featured TCPS after hot

embossing. It is clear that the changes in the surface chemistry, roughness and subsequent wettability of the TCPS surface that result from the hot embossing have a significant effect on the biological response to these substrates. The surface characterisation data presented above indicate that the commercial surface treatment of the TCPS, which increases wettability by the incorporation of oxygen and roughening of the surface [11–13, 22] is degraded by hot embossing. It is thus perhaps not surprising that cell adhesion and proliferation on these degraded surfaces is poorer than for the pristine TCPS surface.

When the different embossed features are considered.

While the wettability of the surface is degraded by the embossing process, as has been discussed, the embossed features appear to provide additional areas for cell attachment on the embossed substrates, and the resulting cell numbers in micro-featured areas are therefore much higher than in non-featured areas. When ridge and pillar features are compared, the shape of the cells and their alignment is significantly different. BAECs cultured on ridge structures preferentially align in the direction of the ridges. Cell alignment has been noted for endothelial cells on micro-patterned topographies [6, 17, 23] and micro-patterned chemical patterns [10, 24, 25] in the literature. Aligned cells on ridge features are also significantly elongated when compared with those cultured on pillar structures or on planar TCPS control surfaces [7, 19, 26, 27]. Cells are typically observed to attach primarily to the tops and edges of the micro-features structures, rather than in the grooves or between pillars. This means that the area available for cellular attachment is higher in the groove direction than in the orthogonal direction, and cells will therefore have a higher probability of aligning in the ridge direction.

The combination of tissue culture treatment and hot embossing is able to provide a mechanism to control both the topography and the surface chemistry of polystyrene cell culture substrates. Spatial control of these surface properties can enable the precise patterning of both adhesive and non-adhesive regions on a single substrate using techniques that are both simple and amenable to industrial application.

4.2.2. Spatial control of tubule formation

After a number of days in culture, specifically on short ridge arrays, BAECs form cord or tube structures, part of the process of angiogenesis. While this

phenomenon has been clearly and repeatably observed after culture on these specific micro-featured surfaces, significant further work is required to understand both the mechanisms involved and the precise nature of these structures.

While angiogenesis has been reported in response to patterns of extracellular matrix [28], polymer fibres [29, 30], biochemical stimuli [31–34], wall shear stress [33, 35], within deep grooves [36, 37] and in co-culture [38–40] the spatial control of tubule formation in response to micro-scale surface features, without biochemical stimulus has not been reported in the literature. This observation has been shown to be repeatable with different cell populations. However, the origin of the mechano-biological stimuli and the biological signaling processes involved are not yet clear [41].

Preferential adhesion to the micro-featured regions is clearly a key step in the processes which enable the spatial control of tubule formation. As has been indicated, preferential adhesion to the micro-featured regions was observed for hot embossed TCPS material, without the use of any biochemical or biological factors.

5. Conclusions

In this paper, we have reported a method that provides for patterning of cell adhesive regions on TCPS, without using biochemical factors. The method produces a surface that has regions of precisely defined micro-structural features with adjoining planar unfeatured regions. The provision of this substrate topography, combined with attendant control of surface chemistry, results in preferential cell adhesion to the micro-featured areas. Alignment is observed, as expected, when endothelial cells are cultured on ridge and groove features, but not on pillar features.

Hot embossing of previously treated Tissue Culture Polystyrene (TCPS) has been shown in this research to have a pronounced effect on both the topography and surface chemistry of the cell culture substrate. This effect that hot embossing has on TCPS surface chemistry has not been reported previously, but clearly has important implications for the subsequent adhesion of cells to these surfaces. An interesting observation that requires further analysis is the formation of cord or tube structures in clearly controlled locations and directions by endothelial cells when cultured on specific patterned substrates.

By using relatively simple fabrication processes, it is therefore possible to create an inexpensive polymer substrate that contains both adhesive and non-adhesive regions. The ability to pattern cell adhesion has been shown previously by adding grafted polymers or protein coatings, but not using a simple bulk processing method as demonstrated here. This methodology may have applications in the design of scaffolds for tissue engineering, and in the development of assays for evaluating angiogenesis, with the potential for use in automated assays for high throughput screening.

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